Phytoconstituent Analysis and Antibacterial Potential of Epicarp Extracts from Mature Fruits of Persea americana Mill

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Background: World Health Organization (WHO) has reported the antimicrobial resistance as one among the ten threats to global health in 2019. The development of plant-derived antibiotics is currently considered as a modern medicine’s greatest success. Persea americana is a plant with high medicinal profile which allow its different parts to be used for therapeutic purposes. This study is aimed to determine the antibacterial potential of ethanol and chloroform extracts from epicarp of mature fruits of P. americana Mill against human pathogens.

Materials and Methods: The epicarps of avocado were dried in oven and ground into powder using porcelain mortar and pestle. The powdered plant materials were extracted with both 96% ethanol and chloroform. Extracts were qualitatively screened to examine their bioactive contents and agar well diffusion method was used to analyze the antibacterial activity of extracts against both Gram-positive and Gram-negative bacteria.

Results: Both solvents showed the ability to dissolve the secondary metabolites from avocado epicarps. Phytochemical screening disclosed the presence of alkaloids, proteins, terpenoids, tannins, flavonoids, steroids and phenolic compounds in ethanolic extracts and absence of flavonoids and tannins in chloroform extracts. The extracts showed the inhibition zones ranging from 14±4.5 mm to 26±2.1 mm while streptomycin demonstrated high inhibition zones ranging from 20±3.1 mm to 30 mm. The minimum inhibitory concentration (MIC) values of extracts ranges from 0.3125 mg/mL to 20 mg/mL while the MIC values for streptomycin vary from 0.25 mg/mL to 1.25 mg/mL.

Conclusion: The ethanol and chloroform extracts proved to be potentially effective as natural alternative preventives to fight against various disease-causing bacteria.

Keywords: antibacterial activity, ethanol extract, chloroform extract, Persea americana, Rwanda
Introduction

Avocado (Persea americana Mill) is a medicinal plant which belongs to the genus Persea of the family Lauraceae. It is believed to have originated from Central America and it is now cultivated in tropical regions worldwide.1 Avocado contains oxygenated carotenoids and the major bioactive compounds that shield human cells from the effects of free radicals. It also consists of persenone A and B which hamper the generation of both superoxide and nitric oxide in cell culture, and fight against inflammation and carcinogenesis.2 Avocado fruit consists of persin which reduces the larval growth of Spodoptera exigua and serve as an antifungal agent against Colletotrichum gloeosporioides.3

Epicarp extracts of avocado is commonly known to have bioactive substances like peptone, alkaloids, steroids, triterpenoids, tannins, flavonoids, saponins, and polyphenols which should have antimicrobial potential against different pathogens.4,5 They also consist of β-galactoside, glycosylated ascorbic acid, cellulose, polygalactourase, polyuronoids, cytochrome P-450, and triacylglycerol which work singlehandedly or collectively to treat monorrhagia, high blood pressure, stomach discomfort, bronchitis, diarrhea and other polygenic disorders including diabetes.1

The unripe avocado fruits contain 1,2,4-trihydroxyhexadec-16-yne which has ability to repress mosquito-borne virus activity without cytotoxicity but by inducing NF-κB-mediated antiviral interferon responses.6 They also have 1,2,4-trihydroxyhexadec-16-ene and 1,2,4-trihydroxyoctadecane which are known to have moderate cytotoxic activities against cancer cells.7,8 Avocado fruits consist of catechins, epicatechins, procanidins and hydroxycinnamic acid with liver repressing activity by modifying the plasma levels of glutamate-pyruvate transaminase, albumin, glutamic oxaloacetic transaminase and creatinine.9,10 Avocado fruits contain vitamins (E and B), potassium and other saturated fatty acids11, which have been documented to possess a variety of protective effects such as antibacterial12, antiviral13, antioxidant14, anti-atherosclerotic15 and hepatoprotective16.

Avocado extracts possess quercetin which has virustatic ability to inhibit human immunodeficiency virus syncytium and viral p24 antigen formations.17 They also contain afzelin and quercetin 3-O-D-arabinopyranoside which have repulsive potential against herpes simplex virus type 1, Aujeszky's disease virus and adenovirus type-3 by inhibiting acyclovir-resistant herpes simplex virus type 1.1

The current research is intended to light its beams on the phytoconstituents screening and antibacterial activity of crude ethanol and chloroform epicarp extracts of mature fruits of P. americana L grown in Rwanda.

Materials and methods

Plant Sample Preparation

Mature ripen P. americana Mill fruits were rigorously collected in Busogo sector, Musanze district, Northern province, Rwanda. Avocado fruits were chosen depending on their solidity, absence of mechanical injury and observable rot. The collected fruits were carefully washed with sterile water and cut open to distinguish the edible parts and epicarps. The epicarps were dried in an oven at 60°C and eventually blended into powder with the help of a porcelain mortar and pestle.

Plant Material Extraction

The extraction experiment was aseptically performed in Biotechnology laboratory at room temperature. P. americana Mill powder was successfully macerated with 96% ethanol (1:5) and chloroform for 3 days using rotary shaker. After extraction, the extracts were decanted and then filtered through Whatman filter paper No. 1. Crude extracts were obtained by evaporating the solvents using rotary evaporator. The yielded thick extracts were dissolved in 2% Sodium Carboxymethyl Cellulose (CMC Na 2%) and kept in labelled containers at 4°C until their use.

Phytochemical Screening

Systematic phytochemical screening was carried out to assess the presence of bioactive components in crude epicarp extracts according to standard method.18 The qualitative analysis tests were performed for various phytoconstituents such as flavonoids (Shinoda test), steroids (Salkowski test), tannins (Ferric chloride test), alkaloids (Wagner test), saponins (Froth test), proteins (Xanthoproteic test) and phenolic compounds were examined by diluting the extract with distilled water up to 5 mL and added 3 drops of 5% ferric chloride solution. The availability of phenols in the extracts was confirmed by the change in colors to dark green. Terpenoids were tested by mixing 5 mL of the crude extracts with 3 mL of chloroform and eventually added 2 mL of concentrated sulphuric acid. The presence
of terpenoids was absolutely revealed by the formation of brown ring.

**Test Bacteria**

In this study, the used bacteria were all human pathogens isolated from clinical specimens. Pure cultures of *Pseudomonas aeruginosa* ATCC 27853, *Staphylococcus pyogenes* ATCC 21059, *Bacillus subtilis* ATCC 6633 and *Klebsiella pneumoniae* ATCC 4352 used to assess the antibacterial properties were obtained from Bacteriology Department in clinical laboratory of University Teaching Hospital of Butare. The bacteria were collected on sterile plates and incubated at 37°C for 24 hours. A single colony of each test bacteria was diluted in 9 mL of peptone water and acclimatized to give the equal concentration of bacterial cells of $10^6$ colony forming unit/mL.

**Antibacterial Assay of The Extracts**

According to National Committee for Clinical Laboratory Standards, the antibacterial activity of the extracts was determined by agar well diffusion method. Twenty-five μL of diluted bacteria were swabbed on agar plates using cotton swabs and Pasteur pipette was used to create wells of 8 mm. Twenty milligrams of each extract (30 mg/mL) was impregnated in created wells and 20 mg of Streptomycin (30 mg/mL) was used as control. The plates were then incubated in upright position at 37°C for 24 hours. All tests were done in triplicate and the antibacterial potential was recorded as the mean±standard deviation by estimating the inhibition zones with Vernier caliper. The activity index for each extract was calculated by dividing the inhibition zone of the sample with the inhibition zone of the standard antibiotic.

**The Determination of Minimum Inhibitory Concentration (MIC)**

MIC was determined by broth dilution method. The extracts were serially diluted from 40 mg/mL to 0.3125 mg/mL and filled in different created wells. Ten μL of each bacterial culture at a density of $10^6$ CFU mL⁻¹ were inoculated on the broth. Control of bacterial growth was also performed and Streptomycin was used as a positive control. All plates were incubated at 37°C for 24 hours. MICs were noted as the smallest concentration of extracts that showed no observable growth of microorganisms after nightlong incubation.

**Results**

**Phytochemical Analysis**

Phytochemical screening clearly confirmed the presence of alkaloids, proteins, terpenoids, tannins, flavonoids, steroids and phenolic compounds in ethanolic extracts and absence of flavonoids and tannins in chloroform extracts. The availability and absence of these secondary metabolites were clearly shown by the results presented in Table 1.

**Antibacterial Activity**

The findings of antimicrobial assay revealed that streptomycin had more potential compared to ethanolic and chloroform extracts. Both extracts were able to inhibit Gram positive bacteria (*S. pyogenes* and *B. subtilis*) stronger than Gram negative bacteria (*P. aeruginosa* and *K. pneumoniae*) (Table 2). This potential was also confirmed by their activity indexes (Table 3) and their respective MICs (Table 4).

The MIC was the lowest concentration that demonstrated the ability to inhibit the visible growth of bacteria after an overnight incubation. From the current findings, streptomycin as a refined antibiotic exhibited high activity compared to both extracts. When the concentration was smaller and its inhibitory activity was high, this confirm

<table>
<thead>
<tr>
<th>Extracts</th>
<th>Flavonoids</th>
<th>Steroids</th>
<th>Terpenoids</th>
<th>Saponins</th>
<th>Tannins</th>
<th>Proteins</th>
<th>Alkaloids</th>
<th>Phenols</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ethanolic</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Chloroform</td>
<td>_</td>
<td>+</td>
<td>+</td>
<td>_</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
</tbody>
</table>

Table 1. The results of the chemical tests of the crude epicarp extracts of *P. americana* fruits.
the high antibacterial activity of tested plant extract. Even if ethanol and chloroform were all organic solvents, ethanol had demonstrated high inhibitory activity than chloroform as it can be observed in Table 4.

### Discussion

This research was conducted to evaluate the phytoconstituents and antipathogenic capabilities of epicarp extracts from fruits of *P. americana* Mill. The main findings of this research revealed that epicarp of avocado contain secondary metabolites with antibacterial properties.

The findings presented in Table 1 disclosed the existence of tannins, steroids, saponins and flavonoids in ethanolic extract which have antibacterial activities. These results are in agreement with the findings reported by other researchers that one or combination of these active substances has/have ability to inhibit microbial cell wall synthesis by creating irremediable compounds with abundant prolene growth factor. The observed plant derived antimicrobials control bacterial growth by modifying their membrane porosity or decreasing their pH. These membrane agitations together with the activity of β-lactams on the transpeptidation of the plasma membrane and eventually accelerate the inhibitory activity of the extracts. The extracts clearly showed the ability to cause leakage of proteins and some enzymes from the cell. These substances have abilities to disrupt binary fission, to interact with extracellular proteins and to damage the integrity of bacterial cell walls. The variability in antipathogenic capabilities of the extracts also depends on the quantity of active substances present in extracted plant parts. Epicarp, roots, leaves, fruits, stems and seeds have different allotment of chemical compounds. The accumulation of the bioactive substances in plant also depends on the stage of maturity, rainfall, seasonality, soil salinity and other agroecological conditions, which impede or enhance water absorption, physiological and chemical processes during plant metabolism.

Both ethanolic and chloroform extracts demonstrated great antibacterial activity against tested bacteria. This result absolutely revealed the effectiveness of organic solvents to dissolve bioactive compounds from plants due to their polarity which influenced their biological activities.

The plant extracts inhibit microbial growth by associating with non-polar compounds in the hydrophobic interior of the membrane and by the formation of hydrogen bonds between the polar head groups of lipids and the more hydrophilic flavonoids at the membrane interface. The antimicrobial activity of flavonoids is explained by the fact that they reduce fluidity in hydrophilic and hydrophobic regions of both inner and outer plasma membrane and cause biofilm perturbation. The high antimicrobial activity of flavonoids is owed to 3-O-octanoyl-epicatechin which enhance membrane affinity of their long acyl chains.

### Table 2. Antimicrobial activity of each epicarp extracts.

<table>
<thead>
<tr>
<th>Bacteria</th>
<th>Inhibition Zones (mm)</th>
<th>Ethanolic Extract (20 mg)</th>
<th>Chloroform Extract (20 mg)</th>
<th>Streptomycin (30 mg/mL)</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>P. aeruginosa</em> ATCC 27853</td>
<td>17±3.3</td>
<td>15±0.9</td>
<td>25</td>
<td></td>
</tr>
<tr>
<td><em>S. pyogenes</em> ATCC 21059</td>
<td>20±0.7</td>
<td>19±0.4</td>
<td>22</td>
<td></td>
</tr>
<tr>
<td><em>B. subtilis</em> ATCC 6633</td>
<td>26±2.1</td>
<td>18±1.6</td>
<td>30</td>
<td></td>
</tr>
<tr>
<td><em>K. pneumoniae</em> ATCC 4352</td>
<td>17±0.2</td>
<td>14±4.5</td>
<td>20</td>
<td></td>
</tr>
</tbody>
</table>

### Table 3. The activity indexes of each epicarp extracts in accordance to the standard antibiotic.

<table>
<thead>
<tr>
<th>Bacteria</th>
<th>Activity Indexes</th>
<th>Ethanol Extracts</th>
<th>Chloroform Extracts</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>P. aeruginosa</em> ATCC 27853</td>
<td>0.68</td>
<td>0.6</td>
<td></td>
</tr>
<tr>
<td><em>S. pyogenes</em> ATCC 21059</td>
<td>0.9</td>
<td>0.86</td>
<td></td>
</tr>
<tr>
<td><em>B. subtilis</em> ATCC 6633</td>
<td>0.86</td>
<td>0.6</td>
<td></td>
</tr>
<tr>
<td><em>K. pneumoniae</em> ATCC 4352</td>
<td>0.85</td>
<td>0.7</td>
<td></td>
</tr>
</tbody>
</table>

Note: Activity indexes=Inhibition zone of extracts/Inhibition zone of standard antibiotic.
Table 4. The MIC of the extracts against the tested pathogens.

<table>
<thead>
<tr>
<th>Bacteria</th>
<th>MIC (mg/mL)</th>
<th>Ethanolic Extract (20 mg)</th>
<th>Chloroform Extract (20 mg)</th>
<th>Streptomycin (30 mg/mL)</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>P. aeruginosa</em> ATCC 27853</td>
<td></td>
<td>5</td>
<td>10</td>
<td>0.75</td>
</tr>
<tr>
<td><em>S. pyogenes</em> ATCC 21059</td>
<td>0.3125</td>
<td></td>
<td>1.25</td>
<td>0.25</td>
</tr>
<tr>
<td><em>B. subtilis</em> ATCC 6633</td>
<td>0.625</td>
<td></td>
<td>2.5</td>
<td>0.5</td>
</tr>
<tr>
<td><em>K. pneumoniae</em> ATCC 4352</td>
<td>10</td>
<td>20</td>
<td>1.25</td>
<td></td>
</tr>
</tbody>
</table>

flavonoids lacking hydroxyl groups on their B rings are the most effective to hinder microbial membranes than those with hydroxyl (OH) groups.34

The findings of the current evaluation showed that Gram-positive bacteria were more sensitive to all extracts than Gram-negative bacteria.35 This statement could be explained by the fact that Gram-positive and Gram-negative bacteria have different cell wall composition. Gram-negative outer membrane consists of phospholipids and lipopolysaccharides that act as a fence to the entrance and reaction of most antibiotics and antimicrobial compounds through cell envelope.22,36 *S. pyogenes* ATCC 21059 as a Gram-positive bacterium showed high sensitivity to the extracts with the activity index of 0.90 as clearly indicated by the findings presented in Table 3. This finding is absolutely correlated with previous result which reported the pronounced sensitivity of *Staphylococcus* species due to their cell walls and outer membranes.37

In the present investigation, antibiotic exhibited high inhibitory activity than the prepared plant extracts as shown in Table 2. The observed effectiveness of antibiotic than the plant extracts may be due to the fact that the antibiotics are in refined states and naturally purified while plant extracts are still in crude states.38

Conclusion

All in all, the present findings provide scientific justification that the epicarp extracts of *P. americana* Mill have potential bioactive substances such as flavonoids and phenolic compounds. From that perspective, avocado extracts contain antimicrobial agents that could be contemplated to develop the effectual treatment modalities for fighting pathogens that are resistant to typical antibiotics. They as well give a great hope that avocado epicarp extracts can be used by pharmaceutical industries to produce valuable medicines to mitigate microbial infections.

References


